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(54) Title: CLONING AND EXPRESSION OF THE EXO-POLYGALACTURONASE GENE FROM *ASPERGILLUS*

(57) Abstract

The present invention discloses a DNA sequence encoding the exo-polygalacturonase gene from *Aspergillus*. Specifically the *Aspergillus tubigensis* exo-polygalacturonase gene is cloned and expressed. The invention relates to vectors comprising the exo-polygalacturonase coding sequence and to host cells transformed with such vectors. The invention further relates to the production of recombinant exo-polygalacturonase and to the use of this protein.

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Cloning and expression of the exo-polygalacturonase gene
from Aspergillus

5

Technical field

The present invention is in the field of molecular
10 biology. Specifically, a DNA sequence encoding the exo-
polygalacturonase gene from Aspergillus is cloned and
expressed. The invention relates to vectors comprising the
exo-polygalacturonase coding sequence and to host cells
transformed with such vectors. The invention further relates
15 to the production of recombinant exo-polygalacturonase.
Furthermore, as an example of the application of this enzyme
partial pectin degradation is demonstrated.

20

Background of the invention

Pectins are major constituents of the cell walls of
edible parts of fruits and vegetables. The middle lamella
which are situated between the cell walls are mainly built up
25 from protopectin which is the insoluble form of pectin.
Pectins are considered as intercellular adhesives and due to
their colloid nature they also have an important function in
the water-regulation of plants. Water-binding capacity is
greatly increased by the amount of hydrophylic hydroxyl and
30 carboxyl groups. The amount of pectin can be very high. For
example, lemon peels are reported to contain pectin up to 30%
of their dry weight, orange peels contain from 15-20% and
apple peels about 10% (Norz, K., 1985. Zucker und Süßwaren
Wirtschaft 38 5-6).

35 Pectins are composed of a rhamno-galacturonan backbone
in which 1,4-linked α -D-galacturonan chains are interrupted
at intervals by the insertion of 1,2-linked α -L-

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rhamnopyranosyl residues (Pilnik, W. and A. Voragen 1970. In 'The Biochemistry of fruits and their products', Vol. 1, Chapter 3, p.53. Acad. Press). Other sugars, such as D-5 galactose, L-arabinose and D-xylose, are present as side chains. A large part of the galacturonan residues is esterified with methyl groups at the C2 and C3 position.

Pectin-degrading enzymes are important tools in the food industry. Traditionally these enzymes are used as 10 mixtures. Aspergillus niger and other fungi produce a whole spectrum of enzymes which can advantageously be used in the degradation of pectin. Examples of such enzymes are pectin esterase, pectin lyase (also called pectin transeliminase), endo- and exo-polygalacturonases. In A. niger the pectin 15 degrading proteins are not expressed constitutively. Induction of these enzymes is achieved by growing the strains when carbon sources such as glucose or sucrose are limiting and in the presence of pectin or breakdown products thereof.

In order to avoid the problem of induction and also to 20 avoid obtaining not well-defined enzyme mixtures there is a growing tendency to clone the genes encoding these enzymes and to express them in other more suitable host cells.

The cloning and expression of several of these enzymes obtained from Aspergillus niger has been reported. 25 EP 0 278 355 describes the cloning of the pectin lyase gene, the sequence thereof and the expression. EP 0 353 188 adds some other pectin lyases.

As mentioned above pectin contains a backbone comprising a high amount of 1,4-linked α -D-galacturonan 30 molecules. This backbone can be digested by the action of depolymerases. Two such depolymerases are known, endo- and exo-polygalacturonase.

Examples of the cloning and expression of especially the former one of these depolymerases have been reported.

35 EP 0 421 919 discloses two polygalacturonases which can be classified as endo-polygalacturonases. Another endo-polygalacturonase has been disclosed in EP 0 388 593.

Both of these patent application used Aspergilli as the source of the gene.

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Exo-polygalacturonases are not so abundant as the endo-form as evidenced by the reports about this enzyme published so far.

Mill (Biochem. J. 99 : 557-561 and 562-565 (1966)) reported the isolation and characterization of two distinct exo-polygalacturonases. One of these was found to be mercury activated. This was confirmed by Hara et al. (Nippon Shokuhim Kogyo Gakkashi 31 : 581-586 (1984)). Kester and Visser (Biotechn. Appl. Biochem. 12 : 150-160 (1990)) report the presence of only one exo-polygalacturonase and 5 endo-polygalacturonases in A. niger culture filtrates. Finally, the cloning of exo-polygalacturonase from Erwinia chrysanthemi EC16 has been reported (He and Collmer J. Bacteriol. 172 : 4988-4995 (1990)).

Exo-polygalacturonase is capable of converting polygalacturonides to galacturonic acid. To enhance this process it is advantageous to use pure exo-polygalacturonase. Galacturonic acid can be used as a source for different synthetic reactions it has for example been found to be useful in the production of ascorbic acid.

Summary of the invention

The present invention provides an isolated recombinant DNA sequence encoding exo-polygalacturonase. Preferably the exo-polygalacturonase is obtained from Aspergilli, more preferably it is obtained from Aspergillus tubigensis or from Aspergillus niger.

The invention also provides an expression cassette containing a DNA sequence encoding the exo-polygalacturonase. Such an expression cassette contains appropriate regulatory regions i.e. a promoter, optionally an enhancer, a terminator. The cassette may also contain a selection marker encoding gene.

In a further aspect of the invention the expression cassette forms part of a vector.

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The invention also provides recombinant host cells transformed with the vector containing the exo-polygalacturonase encoding DNA.

5 The invention further provides isolated recombinant exo-polygalacturonase.

The invention also discloses a method for the production of exo-polygalacturonases. The invention shows how the exo-polygalacturonase can be purified.

10 The invention further discloses a method for obtaining galacturonic acid comprising the use of exo-polygalacturonase.

Finally, the invention describes the use of exo-polygalacturonase in the treatment of pectin.

15

Description of the Figures

20 Figure 1 shows the oligonucleotide mixtures derived from N-terminal amino acid sequences from cyanogen bromide fragments of *A. tubigensis* exo-polygalacturonase PGX. These DNA probes have been used in hybridization experiments to detect the *pgaX* gene.

25 Figure 2 shows the restriction map of the *A. tubigensis pgaX* gene. The open arrow indicates the location and orientation of the gene.

30 Figures 3 and 4 show the results of the use of exo-PG versus Rohament PL for reducing the calcium sensitivity of pectin. X-axis shows molecular weight change (in kD) and Y-axis shows viscosity (in centipoise). The dosages of enzyme in mg per g pectin are printed next to the data points.

o = Rohament PL, + = exo-PG \times = average of blanks.

35

Detailed description of the invention

The present invention discloses purified and isolated
5 DNA sequences which encode a polypeptide having the activity
of the Aspergillus tubigensis PGX enzyme the sequence is
characterized in that it comprises the sequence of SEQ ID
No.1, genetic variants of the sequence of SEQ ID NO 1 are
likewise considered to be part of the invention as are DNA
10 sequences encoding for the same amino acid sequence but
altered making use of the degeneracy of the genetic code.
Further included in the present invention are DNA sequences
capable of hybridizing with the sequence of SEQ ID No. 1 or
with parts thereof.

15 The invention also discloses DNA constructs containing
a DNA sequence encoding PGX wherein the protein encoding
sequence is operably linked to regulatory regions capable of
directing the expression of said DNA sequence in a suitable
host organism.

20 The invention further discloses DNA vectors containing
the DNA constructs encoding PGX.

Transformed microbial host cells containing the vectors
are disclosed and the host cells are shown to produce exo-
polygalacturonase under suitable growth conditions.

25 The present invention discloses a method for obtaining
expression of exo-polygalacturonase comprising, culturing the
microbial host under conditions wherein the cloned gene is
expressed. A method is also disclosed for obtaining exo-
polygalacturonase comprising the steps of culturing the
30 transformed microbial host under conditions giving rise to
the expression of the DNA sequence of and recovering the
polypeptide.

35 The invention also discloses the use of a polypeptide
obtained through the expression of the cloned pgaX gene to
degrade pectin. It is shown in the present invention that the
pectin can be specifically treated using the exo-
polygalacturonase. It is disclosed that this enzyme reduces

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the calcium sensitivity without at the same time reducing the molecular weight of pectin.

Furthermore, the invention provides purified and isolated expression and transcription regulatory regions as found in the 5'non-coding sequences of the Aspergillus tubigensis pgaX gene.

The present invention describes a method for obtaining a gene encoding an Aspergillus exo-polygalacturonase. The 10 method can be summarized as follows.

An Aspergillus strain, which may be selected from the group comprising A. niger, A. awamori and A. tubigensis is grown under conditions wherein the carbon source is limited and wherein pectin or fragments thereof are offered as 15 substrate. These growth conditions give rise to the production of pectin degrading enzymes.

Following sufficient growth time allowing for the accumulation of the different enzymes the cells are harvested and the enzymes are isolated from the medium. After suitable 20 identification the exo-polygalacturonase is purified and sequenced. This sequencing may be performed on the complete protein molecule it may however also be performed on fragments obtained after separation of the peptides following peptidase digestion of the complete protein.

Once the amino acid sequence or parts thereof are determined, suitable sequences are selected for choosing DNA probes. The probes are used against a genomic library of the desired Aspergillus strain. The hybridizing clones are selected and the DNA fragments contained therein are 30 sequenced. The sequence is further completed by adding the rest of the sequence.

Suitable expression regulating sequences are then cloned upstream from the coding region. This depends on the preferred expression host cell. Expression is thus made 35 constitutive.

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The cloned gene is expressed and the medium can be used as such in the degradation of pectin. Preferably, the protein is isolated and used in the desired process.

Alternative ways for cloning, detecting, screening and expressing the gene are known and are expected to give similar results. Among these methods are methods using expression cloning combined with immunological detection, another method would be the use of PCR to amplify the DNA encoding the exo-polygalacturonase gene, this presupposes however that at least a part of the gene sequence is known.

Reaction of the exo-polygalacturonase of the present invention with pectin or pectin-like substrates gives rise to the production of galacturonic acids in monomeric or dimeric form. Galacturonic acid can for example be used in the synthesis of ascorbic acid. Unlike known processes the use of exo-polygalacturonase in pure form gives rise to less by-product formation.

Furthermore, the exo-polygalacturonase of the present invention can be used in combination with pectinesterase, pectinase and other pectin degrading enzymes. It is further shown in the examples that exo-polygalacturonase can be used to treat pectin during this treatment the viscosity of the pectin is reduced without reducing the molecular weight of this polymer.

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Experimental

Throughout this description the following strains and vectors are employed:

5

Strains:

E. coli JM101 (Yanisch-Perron *et al.*, 1985):

thi, Δ (lac-supE, pro AB), [F', traD36, pro AB, lacI^qZ Δ M15]

10

E. coli LE 392 (Murray, 1977):

F' hsdR574 (r_k^+ , m_k^+), sup44, sup58, lacY1, or Δ (lacI^qY)6, galK2, galT22, metB1, trpR55, λ

15

Aspergillus niger N402: cspA1

Aspergillus niger N593: cspA1, pyrA6

Aspergillus tubigensis NW756

Aspergillus tubigensis NW218: pyrA22

Aspergillus nidulans G191: pabaA1, pyrG89, fwA1, uaY9

20

Vectors:

pEMBL18/pEMBL19 (Dente *et al.*, 1983)

25

Example 1

Purification and characterization of Aspergillus tubigensis exo-polygalacturonase PGX

30

Example 1.1

Purification of Aspergillus tubigensis exo-polygalacturonase PGX

35

A culture filtrate (3000 ml) was obtained by the culturing of Aspergillus niger strain NW756 (later

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reclassified as more likely belonging to the species A. tubigensis (Kusters-van Someren *et al.* (1991)) in a medium containing (per liter) 7.5 g NH₄NO₃, 1.5 g KH₂PO₄, 0.5 g KCl, 0.5 g MgSO₄.7H₂O, 2 g yeast extract, trace elements according to Vishniac and Santer (1957) and 10 g degraded polygalacturonic acid as carbon source, pH 6.0.

Polygalacturonic acid was degraded as follows : 40 g polygalacturonic acid (United States Biochemical Corp.) was dissolved in two liters 20 mM sodium acetate buffer pH 4.8 and incubated with 4000 Units of purified A. niger endopolygalacturonase II, of which the purification is described by Kester and Visser (1990), for 24 hours at 30°C.

The pH of the medium was adjusted to 6.0 following inoculation at 10⁶ spores/ml and incubation at 30°C in an orbital shaker at 250 rpm for 55 hours. The culture fluid was passed through cheese cloth and the filtrate was brought to 10 mM sodium acetate, adjusted to pH 4.0 and loaded on a crosslinked alginate column (100 ml) in 20 mM sodium acetate buffer pH 4.0.

The non-bound fraction which contained a mixture of residual endo-PG activity and exo-polygalacturonase activity was concentrated by batch wise adsorption on DEAE Sephadex A-50 (200 g wet weight) at pH 6.0 followed by pulse elution with 1 M NaCl in 20 mM sodium phosphate buffer pH 6.0 and dialysis against the same buffer without the salt. The dialysate was loaded on a DEAE-Sepharose Fast Flow column (2.5 x 10 cm) equilibrated in 10 mM bisTris/HCl buffer pH 5.5.

After elution with a 800 ml linear sodium chloride gradient (0-0.5 M), fractions were assayed for polygalacturonase activity and screened for exo-polygalacturonase activity by a TLC method. This method consists of incubation of samples of the column fractions with 1 % (w/v) polygalacturonic acid in 0.1 M sodium acetate buffer pH 4.2 at 30°C followed by analysis of the reaction products by thin layer chromatography as described by Kester and Visser (1990).

- 10 -

Exo-polygalacturonase positive fractions were pooled, dialyzed against 20 mM sodium acetate buffer pH 4.0 and loaded on a S-Sepharose Fast Flow column (2.5 x 7 cm) equilibrated in the same buffer. Elution was done with a 600 5 ml linear sodium chloride gradient (0-0.5 M).

Final purification was accomplished by repeated chromatography on a MONO Q column (Pharmacia, Uppsala, Sweden) in 20 mM piperazine/HCl buffer pH 6.0. Exo-polygalacturonase containing fractions were diluted five 10 times with water loaded on the column and eluted with a 40 ml linear sodium chloride gradient (0-0.2 M) at a flow rate of 2 ml/minute. A summary of the specific activity and recovery of the enzyme during purification is given in Table I.

15 Table I. Purification of exo-polygalacturonase PGX from
A. tubigensis NW756

20 Step	Total Volume (ml)	Total activity (units)	Specific protein (mg)	activity (units/mg)	Yield (%)
Culture fluid	3000	5230	300	17.4	100
25 Crosslinked alginate	210	3100	212	14.6	59
DEAE-Sepharose	90	846	98	8.6	16
30 S-Sepharose	72	188	8.8	21.4	3.6
MONO Q	10.8	121	3.1	39.0	2.3

35

Polygalacturonase activity was determined by measuring the release of reducing sugars in a reaction mixture containing 0.25 % (w/v) polygalacturonic acid in 0.1 M sodium acetate buffer pH 4.2 at 30°C. Reducing end groups were 40 determined by the neocuproine method described by Stephens *et al.* (1974). One activity unit was defined as the amount of enzyme which produces one μ mole of reducing sugars per minute.

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The apparent molecular mass of exo-polygalacturonase PGX, as determined by SDS-PAGE on a 10 % gel, was 78 kDa. After deglycosylation of the enzyme with N-glycanase (Genzyme, Cambridge, Mass.) according to the supplier 5 instructions the molecular weight was reduced to 52 kDa.

Isoelectric focussing resulted in five distinct bands, all showing exo-polygalacturonase activity, in the pH range 3.7-4.4.

10

Example 1.2

Amino acid sequence determination of the N-termini of cyanogen bromide released peptides of exo-polygalacturonase PGX

15

Approximately 150 µg of exo-polygalacturonase PGX, purified as described in Example 1.1, was extensively dialysed against distilled water, lyophylized and dissolved in 100 µl 70 % (v/v) formic acid-water mixture. To this 20 protein solution 500 µg cyanogen bromide (2500 molar excess), dissolved in the formic acid-water mixture, was added. The reaction was performed for 24 hours at room temperature after which the reaction mixture was diluted 10 fold with water following lyophylization.

25

Approximately one half of the reaction mixture was subjected to electrophoresis on a 15 % polyacrylamide gel, followed by blotting onto Immobilon-P membrane (Millipore) according to the method described by Matsudaira (1987). Membrane slices containing fragments with a molecular mass of 30 kD (CB 1), 27.5 kD (CB 2) and 23.5 kD (CB 3) respectively 30 were used in gas-phase sequencing (Applied Biosystems model 470A protein sequencing, SON, Leiden) according to the program described by Amons (1987).

The following N-terminal amino acid sequences were 35 determined:

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CB 1 (L2278):

1 5 10
5 Ala-(Arg)-Ile-Lys-Val-(Trp)-Pro-Gly-Thr-Pro-Ser-Ala-(Leu)-?-
(Ala)
(Formula 1) = (SEQ ID NO: 1)

10 CB 2 (L1742):

1 5 10
(Gly)-Ile-Ile-Gly-Leu-Asn-Gly-Gly-Thr-Ile-Gly-Pro-Leu-Lys-
15 15 20
Leu-(Arg)-Tyr-Ser-Pro-Gln
(Formula 2) = (SEQ ID NO: 2)

20 CB 3 (L1743):

1 5 10
Phe-Ser-(Leu)-Ser-?-Glu-Ala-Ala-(Thr)-Gly-Pro-Lys-Lys-
(Pro)-Phe-?-? Leu-(Leu)
25
(Formula 3) = (SEQ ID NO: 3)

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Example 2

Screening of the genomic library of Aspergillus niger DS16813 (CBS 323.90; later classified as A. tubigensis) for the exo-polygalacturonase gene (pqaX) and isolation of the gene

5

Example 2.1

Synthesis of a pqaX specific PCR fragment using oligonucleotide mixes derived from PGX amino acid sequences.

10

The amino acid sequences determined in Example 1.2 (Formulas 1, 2 and 3) were used to synthesize oligonucleotide mixes corresponding to the coding and the non-coding DNA strands. The oligonucleotides are synthesized by the 15 phosphoramidite method, using an Applied Biosystems oligonucleotide synthesizer.

The oligonucleotide mixes (Figure 1) were used in PCR with chromosomal DNA of A. tubigensis NW756 as template. All possible combinations of two oligonucleotide mixtures (100 20 pmole each) were used in reactions also containing 0.5 µg chromosomal A. tubigensis NW756 DNA, 200 µM dNTPs, Taq buffer (Sphaero-Q) and 0.5 U Taq polymerase (SuperTaq, Sphaero-Q). Conditions for PCR were 25 cycles of melting at 94°C (1 minute), annealing at 40°C (1 minute) and polymerization at 25 72°C (2 minutes).

The reaction products were analyzed by electrophoresis using a 2% TAE-agarose gel. Only the combination of oligonucleotide mixture 3059 and 3062 resulted in a reaction product, which was approximately 600 bp in length. This PCR fragment was purified using a the Magic PCR Preps DNA purification system (Promega) following the instructions of the manufacturer.

Example 2.2

35

³²P labelling of DNA fragments

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After purification, the DNA concentration of the PCR fragment described in Example 2.1 was determined. 50 ng was labelled by random priming using the Prime-a-Gene kit from Promega. To remove the unincorporated α -32P-dATP from the 5 mixture, the volume was increased to 100 μ l with TE buffer, after which the unincorporated α -32P-dATP was removed by fractionation on a Sephadex G50 column. Fractions containing the radioactively labelled DNA were denatured by incubation for three minutes at 100°C, and kept single stranded by 10 rapidly chilling on ice, before addition to a hybridization buffer containing 6 x SSC, 5 x Denhardt's solution, 0.1% sodium pyrophosphate and 100 μ g/ml heat-denatured herring sperm DNA.

This probe was used in the screening of the genomic 15 library (Example 2.3) and in Southern blot analysis (Examples 2.4, 2.5 and 5.1).

Example 2.3

20 Screening of the Aspergillus tubigensis genomic library for the pqAX gene

To screen for the pqAX gene in an Aspergillus tubigensis genomic library, constructed as described in case 25 EP 0 463 706 A1, Example 2, 5.10^3 pfu per plate were plated in LM top agarose containing 0.7% agarose on 8 85 mm diameter LM (1.2% agar) plates as described (Maniatis *et al.*, 1982, pp. 64). E. coli LE392 were used as plating bacteria.

After incubation of the plates overnight at 37°C, two 30 replicas of each plate were made on nitrocellulose filters (Schleicher and Schüll BA85) as described in Maniatis *et al.* (1982, pp. 320-321).

After baking the filters for 2 hours at 80°C, the filters were prehybridized at 65°C for two hours in a 35 prehybridization buffer containing 6xSSC, 0.5 % SDS, 10xDenhardt's solution and 100 μ g/ml heat denatured herring

- 15 -

sperm DNA (Boehringer). The heat-denatured probe was added and the filters were hybridized for 18 hours at 65°C.

After hybridization, the filters were first washed in 2 x SSC, 0.5% SDS, and then in 0.2 x SSC, 0.5% SDS at 65°C. The 5 air dried filters were taped onto a sheet of Whatman 3MM paper, keying marks were made with radioactive ink and the Whatman paper and filters were covered with Saran Wrap™. Hybridizing plaques were identified by exposure of Kodak XAR X-ray film for 18 hours at -70°C using an intensifying 10 screen.

Twelve plaques hybridizing with the PCR probe, appearing in duplicate on the replica filters, were identified: $\lambda_{\text{pgs}} 1$ to $\lambda_{\text{pgs}} 12$. Each positive plaque was removed from the plate using a Pasteur pipette and the phages were 15 eluted from the agar plug in 1 ml of SM buffer containing 20 μl chloroform, as described in Maniatis *et al.* (1982, pp. 64). The phages obtained were purified by repeating the procedure described above using filter replicas from plates containing 50-100 plaques of the isolated phages.

20 After purification, the phages were propagated by plating 5×10^3 phages on LM medium. After incubation overnight at 37°C, confluent plates were obtained, from which the phages were eluted by adding 5 ml SM buffer and storing the plate for 4 hours at 4°C with intermittent shaking. After 25 collection of the supernatant, the bacteria are removed from the solution by centrifugation at 4,000 x g for 10 minutes at 4°C. Chloroform (0.3%) was added to the supernatant and the number of pfu was determined. These phage stocks contained approximately 10^{10} pfu/ml.

30

Example 2.4

Isolation of DNA from bacteriophage lambda

35 Eight of the isolated phages λ_{pgs} 1, 2, 4, 5, 6, 7, 8 and 11 were propagated as described in Example 2.2 using five plates for each of the phages. The phages were precipitated

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from the thus-obtained supernatant (25 ml) by addition of an equal volume of a solution containing 20% PEG-6000 (w/v) and 2 M NaCl, followed by thorough mixing and incubation on ice for 60 minutes.

5 The precipitated phages were collected by centrifugation at 14,000 x g at 4°C for 20 minutes. The supernatant was removed by aspiration. The phages were carefully resuspended in 4 ml SM buffer and extracted once with chloroform.

10 Prior to extracting the DNA from the phage particles, DNA and RNA originating from the lysed bacteria were removed by incubation of the phage suspension with DNase I and RNase A (both 100 µg/ml) for 30 minutes at 37°C. The phage DNA was subsequently released from the phages by extraction with an 15 equal volume of phenol/chloroform (1:1). After separation of the phases by centrifugation using a Sorvall centrifuge (14,000 x g, 10 minutes), the aqueous phase was extracted once with an equal volume chloroform. The phases were separated by centrifugation (Sorvall centrifuge, 14,000 x g, 20 10 minutes) after which the DNA was precipitated from the aqueous phase by the addition of 0.1 volume 3 M sodiumacetate and 2 volumes ethanol. After mixing, the DNA was recovered by centrifugation in a Sorvall centrifuge for 10 minutes at 4°C (14,000 x g).

25 The supernatant was removed by aspiration after which the DNA was resuspended in 400 µl TE buffer. The DNA was precipitated once again from this solution by the addition of 0.1 volume 3 M sodium acetate and 2 volumes ethanol. The DNA was collected by centrifugation for 10 minutes at 4°C (14,000 30 x g). The supernatant was removed by aspiration, the remaining pellet was briefly dried under vacuum, after which the DNA was resuspended in 125 µl TE buffer containing 0.1 µg/ml RNase A. This purification procedure resulted in the isolation of approximately 10-50 µg DNA from each phage.

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Example 2.5

Restriction analysis of $\lambda_{pq\alpha X}$ containing phages

The isolated DNA of phages $\lambda_{pq\alpha X}$ 1, 2, 4, 5, 6, 7, 8 and 11 was analyzed by Southern analysis using the following restriction enzymes: PvuII and HincII. The DNA was digested for 3 hours at 37°C in a reaction mixture composed of the following solutions; 3 μ l (\approx 1 μ g) DNA solution; 10 μ l of the appropriate 10 x React buffer (BRL); 20 U restriction enzyme (BRL) and sterile distilled water to give a final volume of 100 μ l.

After digestion the DNA was precipitated by the addition of 0.1 volume 3 M NaAc and 2 volumes ethanol. The DNA was collected by centrifugation for 10 minutes at room temperature (14,000 x g). The supernatant was removed by aspiration, the remaining pellet was briefly dried under vacuum and resuspended in sterile distilled water.

After addition of 4 μ l DNA loading buffer the samples were incubated for 10 minutes at 65°C and rapidly cooled on ice. Samples were loaded on a 0.6% agarose gel in TAE buffer. The DNA fragments were separated by electrophoresis at 25 V for 15-18 hours.

After electrophoresis the DNA was transferred to nylon membrane (Gene Bind 45, Pharmacia LKB) by Southern blotting as described in Maniatis *et al.* (1982) and subsequently prehybridized and hybridized using the labeled PCR fragment as described in Example 2.1 and hybridization conditions as described in Example 2.2. The hybridization pattern was obtained by exposure of Kodak XAR-5 X-ray film for 18 hours at -70°C using an intensifying screen.

From the results obtained it was concluded that the DNA of all isolated clones hybridize with the PCR fragment. Two clones appeared to be identical and contained a very large hybridizing PvuII fragment. No hybridizing HincII fragment was detected, suggesting that this fragment was too small and had run off the gel. In six clones fragments originating from the same genomic region were found. In a more extensive

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Southern analysis, using the enzymes XhoI, PvuII, PstI, BglII, SalI, EcoRI and BamHI a partial restriction map of this genomic region was constructed. From this experiment it was concluded that a 5.8 kb XhoI fragment contains the A. tubiqensis pgaX gene.

Example 2.6
Subcloning of the A. tubiqensis pgaX gene

10 From phage λ_{pgax}^7 the 5.8 kb XhoI fragment was isolated by digesting the phage DNA with XhoI and separation of the fragments as described in Example 2.4. The fragment was cut from the agarose gel, after which it was recovered from the 15 agarose using GeneClean (Bio101) as described by the manufacturer. The DNA was dissolved in 10 μ l sterile water and the concentration was determined by agarose electrophoresis, using lambda DNA with a known concentration as a reference and ethidiumbromide staining to detect the DNA.

20 The fragment obtained was ligated in the vector pEMBL19 digested with SalI and dephosphorylated with alkaline phosphatase prepared as follows: 1 μ l (1 μ g/ μ l) pEMBL19 was mixed with 2 μ l 10 x React 10 (BRL), 1 μ l (1 U/ μ l) SalI and 16 μ l sterile distilled water. The DNA was digested for 1 25 hour at 37°C, after which 0.5 μ l alkaline phosphatase (1 U/ μ l) (Pharmacia LKB) was added followed by further incubation at 37°C for another 10 minutes.

The 5.8 kb XhoI fragment was ligated in the vector resulting in the plasmid pIM362, by the following procedure: 30 100 ng pEMBL19 fragment was mixed with 100 ng 5.8 kb XhoI fragment and 4 μ l 5 * ligation buffer (BRL) and 1 μ l (1.2 U/ μ l) DNA ligase (BRL) was added to this mixture in a final volume of 20 μ l. After incubation for 16 hours at 4°C, 10 μ l of the mixture was used to transform E. coli JM101 competent 35 cells, prepared by the CM1, CM2 method as described in the Pharmacia Manual for the M13 cloning/sequencing system. One of the resulting colonies was grown overnight in LB medium

- 19 -

containing 100 μ g/ml ampicillin. From the culture plasmid DNA was isolated by the alkaline lysis method as described by Maniatis *et al.* (1982, pp. 368-369), which was used in restriction analysis, as described in Example 2.4 to check 5 whether it harboured the desired plasmid. Plasmid DNA was isolated on a large scale from 500 ml cultures *E. coli* JM101 containing the plasmid pIM362 grown in LB medium containing 100 μ g/ml ampicillin (Maniatis *et al.*, 1982, p.86) The 10 plasmid was purified by CsCl density gradient centrifugation, phenolized, ethanol precipitated and dissolved in 400 μ l TE. The yield was approximately 500 μ g.

Similarly, a 4.0 kb PvuII fragment which is located within the 5.8 kb XhoI fragment, was isolated from phage 7, subcloned in pEMBL19 digested with SmaI, resulting in plasmid 15 pIM361.

The plasmid pIM361 was further analyzed by restriction enzymes resulting in the restriction map shown in Figure 2.

Plasmids pIM361 and pIM362 containing the exo-PG gene have been deposited in *E.coli* JM101 at the Centraal Bureau 20 voor de Schimmelcultures in Baarn, The Netherlands and have received the deposition number CBS 101.93 and CBS 102.93, respectively.

- 20 -

Example 3

Characterization of the Aspergillus tubigensis pgaX gene

5

Example 3.1

Sequence determination of the A. tubigensis pgaX gene

The sequence of the A. tubigensis pgaX gene was determined by subcloning fragments from pIM362 in pEMBL18/19, 10 in combination with the use of specific oligonucleotides as primers in the sequencing reactions.

For nucleotide sequence analysis restriction fragments were isolated as described in Example 2.5 and then cloned in pEMBL18/19 vectors digested with the appropriate restriction 15 enzymes, as described in Example 2.5. The nucleotide sequences were determined by the dideoxynucleotide chain-termination procedure (Sanger *et al.*, 1977) using the Pharmacia T₇ DNA polymerase sequencing kit. Computer analysis was done using the PC/GENE program (Intelligenetics, Inc.: 20 Madison WI). The sequence was determined for 98% in both orientations Sequence Listing). The sequence includes about 640 bp of the 5' and 655 bp of the 3' sequence.

25

Example 3.2

The A. tubigensis pgaX gene structure

There are probably 6 introns present in the gene. This is based on the appearance of in-frame stopcodons in all of 30 the introns and on the presence of intron consensus sequences (Table II).

Table II. Intron sequences of the pqaX gene.

	position	length	5' site	lariat	3' site
5	Intron I 948-1004	57	GTGctT	ACTAAC	CAG
	Intron II 1372-1423	52	GTGcGT	ACTGAC	TAG
	Intron III 1483-1540	58	GTAAgt	tCTAAC	CAG
	Intron IV 1942-1991	50	GTtaGT	GCTAtC	TAG
	Intron V 2094-2145	52	GTAAgt	gCTGAC	TAG
10	Intron VI 2221-2273	53	GTGAGT	ACTAAC	CAG
	consensus		GTPuNGT	PuCTPuAC	PyAG

15 The pqaX gene encodes a secreted protein of 47.1 kD and an iso-electric point of 4.11. The calculated molecular weight is lower than the one determined by SDS-PAGE (78 kD), even after deglycosylation (52 kD) (there are 12 possible N-glycosylation sites in PGX). Discrepancies between calculated
 20 and determined molecular weights have also been found for other proteins. All three CNBr fragments are found in the sequence. L1743 appears to represent the N-terminal amino acid sequence of the mature exo-PG protein: no methionine is found immediately preceding the phenylalanine (F), which
 25 would have been expected if it were a true CNBr fragment. Instead a sequence with the characteristics of a signal sequence is found. According to the computer program PC-GENE the preferred signal sequence splice site is between serine (S 24) and arginine (R 26), however, since these amino acids
 30 are found in the mature protein, this is not the case. The splice site between the alanine (A 22) and phenylalanine (F 23) is second best.

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Example 4

Expression of the pqax gene in Aspergillus niger N593, Aspergillus tubigensis NW218 and Aspergillus nidulans G191

5

Example 4.1

Introduction of the pqax gene into Aspergillus niger N593, Aspergillus tubigensis NW218 and Aspergillus nidulans G191 by cotransformation

10

The plasmids pIM361 and pIM362, obtained in Example 2.5, were introduced in A. niger, A. tubigensis and A. nidulans by cotransformation of A. niger N593, A. tubigensis NW218 and A. nidulans G191 using the A. niger pyrA gene as a 15 selective marker on the plasmid pGW635 (Goosen *et al.*, 1989) and the plasmids pIM361 and pIM362 as the cotransforming plasmids in separate experiments.

Protoplasts were prepared from mycelium by growing A. niger N593 and A. tubigensis NW218 on minimal medium 20 supplemented with 0.5% yeast extract, 0.2% casamino acids, 50 mM glucose and 10 mM uridine for 20 hours at 30°C. For the preparation of protoplasts A. nidulans G191 was grown at 37°C on the same medium supplemented with 1.35 mg/l p-aminobenzoate. The preparation of protoplasts and the 25 transformation procedures were performed as described by Goosen *et al.*, 1987. The resulting pyr' transformants were then analyzed for the expression of the pqax gene by Western blot analysis.

30

Example 4.2

Screening of transformants for the expression of the pqax gene

35 The transformants obtained in Example 4.1 were analyzed for the formation of the pqax gene product, the PGX protein. 10 pIM361 and 10 pIM362 A. niger transformants, 5 pIM361 and

- 23 -

5 pIM362 A. tubigensis transformants and 10 pIM361 and 10 pIM362 A. nidulans transformants were selected and grown for 24 hours on medium containing per liter 10 g PGII-degraded polygalacturonic acid, 7.5 g NH_4NO_3 , 0.5 g KCl, 0.5 g 5 $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$, 1.5 g KH_2PO_4 , 0.2% yeast extract, 1 ml/l Vishniac solution and 1.35 mg/l p-aminobenzoate for A. nidulans only (pH 6.0).

After growth (24 hours after inoculation) the mycelium was removed by filtration and the proteins in the culture 10 filtrates were analyzed by SDS-polyacrylamide gel electrophoresis, using a gel containing 10% acrylamide.

The PGX protein was detected on nitrocellulose after electroblotting and incubation with polyclonal antibodies raised against the PGX protein purified as described in 15 Example 1.1. The antibody bound, was detected after incubation with goat-anti-mouse antibody conjugated to alkaline phosphatase, according to the Biorad instruction manual.

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Table III. Transformants which overproduce PGX.

t=24		
5	<u>A. tubigensis</u> (pIM361)4	++
	(pIM361)6	+++
	(pIM362)7	+
10		
	<u>A. nidulans</u> (pIM361)6	++
	(pIM361)9	+++
15	(pIM362)2	++

The amount of PGX produced as judged by Western analysis is indicated.

20 +++: high overproduction as compared to the corresponding wild type strain.

+ : low overproduction as compared to the corresponding wild type strain.

25 Table III shows the results of the Western analysis. Only A. tubigensis and A. nidulans transformants readily overproduced the PGX protein as detected by this procedure. The protein was secreted into the medium. Of the A. tubigensis transformants analyzed one was selected for giving the highest yields of the PGX protein, transformant A. tubigensis NW218(pIM361)6.

In a second experiment A. tubigensis NW218 was transformed with 1 μ g of pGW613 which contains the A. niger 35 pyrA gene and 40 μ g pIM361. The highest producer found in this experiment was NW218 (pIM361)22.

- 25 -

Example 4.3

Overexpression of PGX in *A. tubigensis* transformants and purification and characterization of PGX

PGX was purified from *A. tubigensis* NW756. NW756 was grown for 48 h at 30°C in medium containing 7.5 g ammonium nitrate, 1.5 g KH_2PO_4 , 0.5 g KCl, 0.5 g $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$, 2 g yeast extract, trace elements and 1% PGII-degraded polygalacturonic acid, pH 6.0. The culture fluid was harvested by filtration and the pH (initial value 6.8) was adjusted to 6.0. The culture filtrate was stirred for one hour with DEAE-Sephadex A50 (300 g, wet weight), equilibrated in 10 mM sodium phosphate buffer pH 6.0. The ion-exchanger was collected, poured into a column, and bound protein was eluted by a pulse of 1 M NaCl in buffer. After dialysis against 20 mM sodium acetate buffer pH 4.2 the solution was loaded on a cross-linked alginate column ($0=5$ cm, $h=8$ cm) in the same buffer followed by elution with two pulses of respectively 0.3 and 1 M NaCl in buffer. Part of the exo-PG activity did bind to the crosslinking alginate column and was eluted by the 0.3 M NaCl pulse (pool I). The non-bound fraction was dialyzed against 20 mM sodium citrate buffer pH 3.5 and loaded on a S-Sepharose Fast Flow column ($0=2.5$ cm, $h=25$ cm) equilibrated in the same buffer followed by elution by a 600 ml NaCl gradient (0-0.6 M) (pools I, II, III). These pools all contained exo-PG activity and all also contained endo-PG activity except pool I of the S-Sepharose column.

All four pools were concentrated up to 5 ml by adsorption on a small ($0=1.5$ cm, $h=5$ cm) DEAE-Sepharose Fast Flow column at pH 6.0 followed by pulse elution with 1 M NaCl in 10 mM sodium phosphate buffer pH 6.0. The final purification was done by GPC on a Sephacryl-S 200 column ($0=2.5$ cm, $h=90$ cm) in 0.1 M sodium acetate buffer pH 4.8.

No endo-PG and PL activity could be detected in these final samples.

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	volume	U/ml	mg/ml	S.A.
crl. gen. pool I/GPC/conc.	9.8	19.4	0.54	36
S-Seph. pool II/GPC	22.7	1.28	0.017	75
S-Seph. pool III/GPC/conc.	6.5	14.4	0.31	46.5

5 S.A. = specific activity

Purification of PGX from transformant NW218 (pIM361) 22.

10 Cultivation was done for 50 h at 30°C in minimal medium with ammonium nitrate as nitrogen source and 1% PGII-degraded polygalacturonic acid as carbon source. After 24 and 50 h only one major protein could be detected in the culture fluid as judged by Coomassie Brilliant Blue-staining following SDS-polyacrylamide gel electrophoresis. Total protein in the 15 culture filtrate was concentrated by batch wise adsorption on DEAE A50 at pH 6.0 followed by pulse elution with 1 M NaCl in 20 mM sodium phosphate buffer pH 6.0.

20 After extensive dialysis against 20 mM sodium acetate buffer pH 3.6 dialysate was loaded onto a S-Sepharose Fast Flow column (2.5 * 20 cm) equilibrated in the same buffer. Elution was done with a linear NaCl gradient (800 ml, 0 - 0.5 M).

25 Fractions were screened by SDS-PAGE and based on these results two pools were formed which were used for further analysis.

	U/ml	mg/ml	U/mg	mg	total U
Pool I	39.5	0.87	45.4	61	2.765
Pool II	400	5.1	78	790	12.900

30

Based on these results the production level of PGX in this transformant (approx. 180 mg/l) is 80 fold higher than for the wild type strain NW756.

35 After one purification step the preparation is not completely pure, but no pectin lyase and no endo-PG activity

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could be detected. Pool I contains a low level of pectinesterase activity.

5

Example 4.4

Deregulated expression of the A. tubigensis pgaX gene

To make a gene fusion of the A. niger N400 pki promoter and the A. tubigensis DS16813 pgaX gene, first a PCR fragment 10 was made using a primer which contains the pki SmaI site and another primer containing the A. niger pki ATG and the first 12 nucleotides of the pgaX gene (pgaX sequences are shown in small letters):

15

<u>pki SmaI</u>	5'	3'	<u>pki-pgaX</u>	5'
TTTCCTT <u>CCCGGG</u> CAC			GTTAGTAGGCAGTTCTACtctgagtgcgtg	
sense			antisense	
20	SEQ ID NO: 6		SEQ ID NO: 7	

25

Similarly, a PCR fragment was made using an oligo complementary to the pki-pgaX primer described above and another primer containing a NcoI site 165 bp upstream in the pgaX coding region:

30

<u>pki-pgaX</u>	5'	3'	<u>pgaX NcoI</u>	5'
CAATCATCCGTCAAGATGagactcacgcac			cggtagaccttaccgtga	
sense			antisense	
35	SEQ ID NO: 8		SEQ ID NO: 9	

40

Subsequently, both PCR fragments were used in a third PCR experiment with only the pki SmaI primer and the pgaX NcoI primer. This resulted in a fragment of approx. 300 bp which was digested with SmaI and NcoI to obtain the desired sticky ends and the fragment was then further purified.

45

pIM361 was digested with BamHI which cuts in the pEMBL polylinker. A BamHI fragment containing the pki promoter and

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the A. niger pelB gene was cloned into this plasmid. Digestion with NcoI and SmaI cut out the pelB gene as well as the 5' end of the pgaX gene and the 3' end of the pki promoter. Ligation of the SmaI-NcoI PCR fragment described 5 above resulted in pIM365 which thus contains the A. niger pki promoter fused to the A. tubigensis pgaX gene. The construct was checked by sequencing, using the commercial universal and reverse primers as well as the pki-SmaI and pgaX-NcoI primers to sequence the PCR fragment.

10 A. tubigensis NW218 was transformed with the A. niger pvrA gene (pGW635, 1 μ g) and pIM365 (40 μ g) as cotransforming plasmid. Only two PGX overproducing transformants have been found, NW218 (pIM365) number 52 and 59, of which 59 has the highest production level. The copynumber of pgaX in this 15 transformant has not yet been established.

Example 5

Screening for genes related to the pgaX gene in Aspergillus 20 niger, Aspergillus tubigensis and Aspergillus nidulans

Example 5.1

Genomic hybridization of A. niger, A. tubigensis and A. 25 nidulans DNA

High molecular weight DNA isolated from A. niger N400, A. tubigensis NW756 and A. nidulans WG312 as described in the published european patent application EP 0 463 706, Example 30 2.1, was digested with EcoRI, BamHI and HindIII. The resulting fragments were separated by agarose gel electrophoresis and transferred to nitrocellulose membrane as described by Maniatis et al. (1982, pp. 383-389).

The nitrocellulose membranes were prehybridized at 60°C 35 for two hours in hybridization buffer (as described in Example 2.2, above). After prehybridization, the radioactively labelled fragment, described in Example 2.2,

- 29 -

was added to the hybridization buffer and the hybridization was continued for 18 hours. After hybridization, the filters were washed for 60 minutes at 60°C in 4 x SSC, 0.5% SDS followed by final washing using 2 x SSC, 0.5% SDS at 60°C.

5 After taping the membranes to Whatmann 3MM paper and properly marking with radioactively labelled ink, the filters were covered with Saran Wrap™ and autoradiographed for 72 hours at -70°C using Kodak XAR-5 X-Ray film and Kodak X-Omatic cassettes with regular intensifying screens.

10 The hybridization fragments found are summarized in Table IV.

Table IV. Hybridizing fragments and their lengths (kb) found
15 in A. niger, A. tubigensis and A. nidulans genomic DNA using
a fragment of the A. tubigensis pqaX gene as a probe.

		<u>Eco</u> RI	<u>Bam</u> HI	<u>Hind</u> III
20	<u>A. niger</u> N400	13	9.0 6.0 4.4*	2.5 1.9 1.6*
25	<u>A. tubigensis</u> NW756	5.5	23 ¹ 12 ¹ 9.0*	2.5
30	<u>A. nidulans</u> WG312	4.4	1.3	3.4

* : strongest hybridizing fragment

¹ : partially digested

- 30 -

Example 6

Effect of exo-polygalacturonase on calcium sensitivity and molecular weight of pectin

5 Methods

Exo-PG and Rohament PL were used in these experiments.

Exo-PG was isolated from the supernatant of an *A. tubigensis* culture. Exo-PG was stored frozen in 20mM sodium citrate buffer pH 3.5. Exo-PG had an activity of 14.4 U/ml and a concentration of 0.31 mg/ml. No lyase and endo-polygalacturonase activities could be detected in the exo-PG solution. The activity was measured on polygalacturonic acid at a concentration of 0.25% (w/v) in 0.1 M sodium acetate pH 4.2 at 30°C.

15 Rohament PL was obtained from Rohm GmbH (Darmstadt, Germany). The activity was 2500 pgu/ml measured according to internal Rohm standards.

Pectin used for the test system has the following characteristics:

20 degree of ester = 71.7 (by titrimetry),
molecular weight = 117 kD, as described below,
calcium sensitivity = 213, according to the test described below.

25 Molecular weight determination for pectin

Principle

Molecular weight is estimated by measuring the relative viscosity of a 0.1% pectin solution using Na-hexametaphosphate.

30

Apparatus

1. Ostwald capillary tube viscosimeters (no less than two) with 100 to 150 sec. outlet time for water (25°C). For instance Silber Brand # 111.
- 35 2. Transparent thermostated water bath, 25.0°C ± 0.3°C.

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Reagents

1. Na-hexametaphosphate solution:
 - a) 20.0 g Na-hexametaphosphate is dissolved in 1800 ml ion exchanged deaerated (boiled) water.
 - 5 b) pH is adjusted to 4.50 ± 0.05 with 1 M HCl.
 - c) The solution is diluted with ion exchanged, deaerated (boiled) water until 2000 ml.

Procedure

- 10 1. The viscosimeters must be cleaned appropriately.
2. Outlet time for hexametaphosphate solution is measured (section: Measuring of outlet time) on the viscosimeters used every time a new hexameta-phosphate solution is prepared and for every new working day where pectin 15 solutions are being measured.
Immediately before measuring the necessary quantity of hexametaphosphate solution is filtered through a glass filter # 3.
- 20 3. The pectin sample system for molecular weight determination is made as follows:
 - a) Acid wash the pectin as described in the method for determination of AGA and DE (Food Chemicals Codex, 3rd Edition, Washington D.C., 1981).
 - 25 b) Approx. 90 g hexametaphosphate solution is weighed in a tared beaker with magnet.
 - c) 0.1000 g acid washed pectin is gradually added while stirring. Keep stirring until the pectin is completely dissolved.
 - d) Weigh up to 100.0 g with hexametaphosphate solution.
 - 30 e) Filter through a glass filter x 3.
4. For every molecular weight determination the outlet time is measured (section: Measuring the outlet time) for the pectin/hexametaphosphate solution on two different viscosimeters.
- 35 5. Molecular weight is calculated (section: Calculation) separately for each viscosimeter using the latest

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measured outlet time for hexametaphosphate solution on the viscosimeter in question.

6. Should the difference between two calculated molecular weights be less than 3500 the mean value is calculated.
- 5 Round off the value to the nearest multiple of 1000 and that will be the result of the method.
7. Should the difference between the two calculated molecular weights be 3500 or more the viscosimeters should be cleaned and a new measuring of outlet time for
- 10 hexametaphosphate solution should be performed.

Measuring the outlet time

1. The viscosimeter is rinsed twice with the sample.
2. Pour 5.00 ml of the sample in the viscosimeter and place it in the thermostated water bath at $25.0^{\circ}\text{C} \pm 0.3^{\circ}\text{C}$ at least 15 minutes prior to measuring.
- 15 3. Time is measured on two outlets. Should the difference between the times be more than x seconds the measuring is repeated until you have three outlet times which differ no more than x seconds.
- 20 x = 0.2 seconds on measuring hexametaphosphate solution
x = 0.4 seconds on measuring samples
4. The outlet time which is needed for further calculations is the mean value of the above mentioned two or three
- 25 identical or almost identical measuring results.

Calculation

The relative viscosity is calculated, as follows:

$$30 \quad n_r = \frac{\frac{K}{(t_o - t_o)}}{\frac{K}{(t_h - t_h)}}$$

35

Where t_o and t_h are outlet times for pectin solution and hexametaphosphate solution, respectively.

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The parameter K can with sufficient accuracy be fixed at 75 s² using Silber Brand No. 111 viscosimeter. Otherwise, K can be calculated as follows:

5

$$K = \frac{Q \times t_v^2}{Q + (0.226 \cdot L \cdot t_v)}$$

10 where Q = volume of viscosimeter bulb in cm³, L = length of capillary tube in cm and t_v outlet time for water in seconds.

The molecular weight of pectin is calculated as follows:

15

$$M = \frac{(n_r^{1/P} - 1) \cdot P}{k \cdot C}$$

20

where P is fixed at 6 and k is fixed at 4.7 · 10⁻⁵ mol · g⁻¹; C is the weight percentage of pectin in the sample system - i.e. 0.1% with the numerical values inserted you will obtain:

25 M = 1.277 · 10⁶ (n_r^{1/6-1}) g/mol.

Literature

P.E Christensen (1954) and Smit and Bryant (1967)

30

Calcium sensitivity test

A calcium containing pectin solution is prepared under 35 conditions avoiding the formation of local gels during the addition of the calcium salt to the pectin solution.

The calcium salt and pectin are mixed at low pH prohibiting the reaction between calcium and pectin. This reaction is initiated by increasing the pH by adding an 40 acetate buffer to obtain the desired pH.

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Reagents:

1 M HCl
1 M Acetate buffer, pH 4.75
250 mM Calcium chloride solution

5

A pectin solution of the desired concentration is prepared in distilled water, and pH adjusted to 1.5 with 1 M HCl.

10 145 g portions of this pectin solution are measured into viscosity glasses.

5 ml of the calcium chloride solution is added to the 145 g pectin solution to give a final concentration of 8.3 mM Ca.

15 With efficient stirring with a magnetic stirrer, 25 ml of the acetate buffer is added to the pectin solution to bring pH to 4.2.

The magnet is taken out, and the glass is left at room temperature (25°C) until the next day, when the viscosity is measured with a Brookfield viscosimeter.

20 The pectin used in the present experiments had calcium sensitivity = 213, according to this test.

Test for polygalacturonase activity

25 26 Two enzymes are tested in every experiment: exo-PG and Rohament PL.

30 (1) A 1.6% pectin solution is prepared, and pH is adjusted to 3.8 with dilute NH₃ solution or dilute HCl. The solution is kept at 38°C.

35 (2) Three dilutions of each enzyme (in 20 mM sodium citrate buffer, pH 3.5) are prepared. The concentrations are chosen in accordance with the expected activity of the enzymes so a considerable (but not exhaustive) reduction of the calcium sensitivity of the pectin is achieved when the test system is incubated with the enzyme dilutions as will be detailed in the following

- 35 -

text. Further, the concentrations of the three dilutions are in the relative proportions 1 : 2 : 4.

(3) The pectin solution is split in nine portions, 95 g each. Three of these are incubated with 5 ml of each dilution of the "unknown" enzyme. Three portions are incubated with the "standard" enzyme. The remaining three portions are incubated with 5 ml water, they are referred to as "blanks".

(4) All nine test systems are treated in the same way:

- (a) incubation at 38°C for 20 hours
- (b) addition of 100 ml diluted HCl, the concentration is chosen by experience so that the pH of the test system becomes 1.8 ± 0.2
- (c) heating on water bath for 4 minutes at 85°C, measured from the time when 70°C was reached (this treatment should destroy the enzyme)
- (d) cooling to 25°C on water bath.

(5) Each of the so treated pectin solutions is split into two portions: one of these is made into - and measured as - a test system for pectin molecular weight, while the other is made into a test system for calcium sensitivity.

The test system for molecular weight is 0.1% pectin in sodium hexametaphosphate solution, pH 4.5. The viscosity is measured in an Ostwald capillary and converted to (what is reported as) molecular weight by a numerical expression. Details are given above

The test system for calcium sensitivity is 0.6% pectin in sodium acetate buffer with added Ca^{++} , pH 4.4. The viscosity is measured by means of a Brookfield viscosimeter. Details may be found above.

Results

The results are reported in a diagram with MW-change (in kD, relative to the average of the blanks) as the x-axis and calcium sensitivity (in cp) as the y-axis. An enzyme showing desired performance, i.e. a low calcium sensitivity

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is achieved with almost no loss in molecular weight, will have data points in the lower right corner of the diagram.

Results of the two independent experiments are shown as Figures 3 and 4. The dosages of enzyme in mg per g pectin are 5 printed next to the data points.

The Figures presented lead to the conclusion that:

- (1) The exo-PG preparations can reduce the calcium sensitivity of the pectin that was used in the test 10 system
- (2) In both experiments, data points for the exo-PGs tend to be to the lower right side of the data points for the standard (Rohament PL) in the diagrams, Figures 3 and 4. This suggests that for this particular purpose 15 an exo-PG is more desirable in its way of action than Rohament PL.

The above data suggest that a great part of the calcium sensitivity of pectin is due to blocks situated at the non-reducing ends of some of the pectin molecules. 20

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SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT:

- (A) NAME: Gist-brocades
- (B) STREET: Wateringseweg 1
- (C) CITY: Delft
- (E) COUNTRY: The Netherlands
- (F) POSTAL CODE (ZIP): 2611 XT

(ii) TITLE OF INVENTION: Cloning and expression of the
exo-polygalacturonase gene from *Aspergillus*

(iii) NUMBER OF SEQUENCES: 15

(iv) COMPUTER READABLE FORM:

- (A) MEDIUM TYPE: Floppy disk
- (B) COMPUTER: IBM PC compatible
- (C) OPERATING SYSTEM: PC-DOS/MS-DOS
- (D) SOFTWARE: PatentIn Release #1.0, Version #1.25 (EPO)

(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 12 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(v) FRAGMENT TYPE: internal

(vi) ORIGINAL SOURCE:

- (C) INDIVIDUAL ISOLATE: L2278

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

Ala Xaa Ile Lys Val Xaa Pro Gly Thr Pro Ser Ala
1 5 10

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 19 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

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- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iii) ANTI-SENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: L1742

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

Ile Ile Gly Leu Asn Gly Gly Thr Ile Gly Pro Leu Lys Leu Xaa Tyr
1 5 10 15
Ser Pro Gln

(2) INFORMATION FOR SEQ ID NO: 3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 18 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: L1743

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

Phe Ser Xaa Ser Xaa Glu Ala Ala Xaa Gly Pro Lys Lys Xaa Phe Xaa
1 5 10 15
Xaa Leu

(2) INFORMATION FOR SEQ ID NO: 4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2974 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)

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(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Aspergillus tubigensis
(B) STRAIN: NW756

(ix) FEATURE:

(A) NAME/KEY: exon
(B) LOCATION: 640..946

(ix) FEATURE:

(A) NAME/KEY: intron
(B) LOCATION: 947..1003

(ix) FEATURE:

(A) NAME/KEY: exon
(B) LOCATION: 1004..1370

(ix) FEATURE:

(A) NAME/KEY: intron
(B) LOCATION: 1371..1422

(ix) FEATURE:

(A) NAME/KEY: exon
(B) LOCATION: 1423..1481

(ix) FEATURE:

(A) NAME/KEY: intron
(B) LOCATION: 1482..1539

(ix) FEATURE:

(A) NAME/KEY: exon
(B) LOCATION: 1540..1940

(ix) FEATURE:

(A) NAME/KEY: intron
(B) LOCATION: 1941..1990

(ix) FEATURE:

(A) NAME/KEY: exon
(B) LOCATION: 1991..2092

(ix) FEATURE:

(A) NAME/KEY: intron
(B) LOCATION: 2093..2144

(ix) FEATURE:

(A) NAME/KEY: exon
(B) LOCATION: 2145..2219

(ix) FEATURE:

(A) NAME/KEY: intron
(B) LOCATION: 2220..2272

(ix) FEATURE:

(A) NAME/KEY: exon

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(B) LOCATION: 2273..2320

(ix) FEATURE:

(A) NAME/KEY: CDS
 (B) LOCATION: join(640..946, 1004..1370, 1423..1481,
 1540..1940, 1991..2092, 2145..2219,
 2273..2320)
 (D) OTHER INFORMATION: /codon_start= 640
 /product= "exo-polygalacturonase precursor"
 /gene= "pgalX"

(ix) FEATURE:

(A) NAME/KEY: sig_peptide
 (B) LOCATION: 640..705

(ix) FEATURE:

(A) NAME/KEY: mat_peptide
 (B) LOCATION: 706..2320
 (C) IDENTIFICATION METHOD: experimental
 (D) OTHER INFORMATION: /product= "exo-polygalacturonase
 mature enzyme"
 /evidence= EXPERIMENTAL

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

GA	CTCTAGAG	GATCCCCCTG	TGGCCCTTTT	AAATGGCTTC	CTAGTGGTGG	TGTATGTCC	60
AC	GGGGCTTG	TACCAACACGG	AGTTTGCCT	GACCTGCACT	TACCGCAAGT	TTCGGGAATT	120
GC	CTGCGCG	ACAATATGAT	CTAACGGGGA	TTAAAGAATA	ACCGGGTATT	GCTCTCTCAA	180
TT	AAAATGAA	ATCATGATGC	AGAAGATGGC	TGGGGTAAGT	TCCCTGCCAA	TTCCCCAGAA	240
AA	ACACGTTTTT	CCCCCAATCG	GATAATGTC	TCCGACAAAGC	CCTCCATATCA	CATCGCTCT	300
CC	GGTGCCT	CCACGGAAATC	CCCCACCAACC	ACATTCCTCCA	GGAGCTAACT	GTCTCCAAGT	360
GG	CCCCTGTTG	ATTACAGGAG	CAAGCCCAA	GCTTACGATA	GCTCCACAGC	CGTGGTGTGC	420
CC	TCACTCCGTT	TGGTCGATTT	TGTTTCCCG	GATATGGAGT	CCAGGCAACT	CCCCAACTCG	480
TT	GAGACATGGG	TGGTGIGGGC	AGTGTGAGCA	AGACTGCCCCG	GGAGGCAATGT	CGACTCAGTG	540
TT	CAATGGATAT	AAATGCCAGT	GGGAGATGTC	GTTCCTCTGC	GGTAATCCTG	CCCAAGCTCT	600
CC	GTCCGGAAAC	CCAGGACTTGT	CTCCCTGCCAA	TTACACCAA	ATG AGA CTC ACG CAC		654
AA					Met Arg Leu Thr His		
					-22	-20	
GT	CTT CTC TCG	CAC ACG CTT	GGC CTT CTT	GCG CTA GGG	GCC ACA GCA	GGG	702
TT	Val Leu Ser	His Thr Leu	Gly Leu	Leu Ala	Leu Gly	Ala Thr	
CC	-15	-10	-5				
TT	GCC TTC TCC	CGA TCC AGA	GAA GCT GCG	TGC GGC CCA	AAA AAG CCT	TTC	750
CC	Ala Phe Ser	Arg Ser Arg	Glu Ala	Ala Cys	Gly Pro Lys	Lys Pro Phe	
AA	1	5	10	15			

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CGG CCT CTA CCT ACA AGC CAG AGC AGG GAC AAG ACC TGC CAT GTC CGC Arg Pro Leu Pro Thr Ser Gln Ser Arg Asp Lys Thr Cys His Val Arg 20	25	30	798
AGC CAT GGA GAT GGC ACT GAC GAC TCT GAT TAC ATT CTC TCC GCA TTG Ser His Gly Asp Gly Thr Asp Asp Ser Asp Tyr Ile Leu Ser Ala Leu 35	40	45	846
AAC CAA TGT AAC CAC GGT GGA AAG GTA GTT TTT GAT GAG GAC AAG GAA Asn Gln Cys Asn His Gly Gly Lys Val Val Phe Asp Glu Asp Lys Glu 50	55	60	894
TAC ATT ATC GGG ACG GCA CTG AAT ATG ACC TTC CTG AAG AAC ATT GAC Tyr Ile Ile Gly Thr Ala Leu Asn Met Thr Phe Leu Lys Asn Ile Asp 65	70	75	942
CTA G GTGCTTATTC TGCAGACCCA ATCAAGGTGA CCATTGACTA ACCTTATGGT Leu 80			996
TCAACAG AG GTC CTC GGA ACA ATC TTA TTC ACT AAC GAT ACA GAC TAC Glu Val Leu Gly Thr Ile Leu Phe Thr Asn Asp Thr Asp Tyr 85	90		1044
TGG CAA GCC AAC TCC TTC AAA CAG GGC TTC CAG AAC GCT ACG ACC TTC Trp Gln Ala Asn Ser Phe Lys Gln Gly Phe Gln Asn Ala Thr Thr Phe 95	100	105	1092
TTC CAA CTC GGT GGT GAA GAT GTG AAT ATG TAC GGT GGT GGT ACA ATC Phe Gln Leu Gly Gly Glu Asp Val Asn Met Tyr Gly Gly Thr Ile 115	120	125	1140
AAT GGC AAC GGA CAG GTC TGG TAT GAT CTG TAT GCC GAA GAT GAT CTC Asn Gly Asn Gln Val Trp Tyr Asp Leu Tyr Ala Glu Asp Asp Leu 130	135	140	1188
ATT CTG CGT CCC ATC TTG ATG GGC ATC ATT GGG CTG AAT GGT GGC ACA Ile Leu Arg Pro Ile Leu Met Gly Ile Ile Gly Leu Asn Gly Thr 145	150	155	1236
ATT GGT CCG CTG AAG CTG CGG TAC TCG CCG CAA TAC TAC CAT TTT GTG Ile Gly Pro Leu Lys Leu Arg Tyr Ser Pro Gln Tyr Tyr His Phe Val 160	165	170	1284
GCT AAC TCG TCG AAT GTG CTC TTT GAC GGG ATT GAC ATT TCG GGT TAT Ala Asn Ser Ser Asn Val Leu Phe Asp Gly Ile Asp Ile Ser Gly Tyr 175	180	185	1332
AGT AAG AGC AAC GAA GCC AAA AAC ACT GAT GGA TG GTGCGTTTA Ser Lys Ser Asp Asn Glu Ala Lys Asn Thr Asp Gly Trp 195	200		1380
TCCTGCTTTA CACTGAGCGT TATACTGACC TTTTCCCGT AG G GAT ACC TAC CGC Asp Thr Tyr Arg 205			1435

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TCG AAC AAT ATC GTT ATC CAG AAT TCG GTG ATC AAC AAC GGT GAT G	1481
Ser Asn Asn Ile Val Ile Gln Asn Ser Val Ile Asn Asn Gly Asp	
210 215 220	
GTAAAGTAAA CCTAAGTAGC GTCATACITC AACATTCTA ACCTGAAAC CTACACAG AC 1541	
Asp	
TGT GTC TCT TTC AAG CCG AAC AGC ACC AAT ATC CTC GTT CAG AAC CTT 1589	
Cys Val Ser Phe Lys Pro Asn Ser Thr Asn Ile Leu Val Gln Asn Leu	
225 230 235	
CAC TGC AAT GGC TCC CAC GGC ATT TCT GTT GGC TCT CTC GGC CAA TAC 1637	
His Cys Asn Gly Ser His Gly Ile Ser Val Gly Ser Leu Gly Gln Tyr	
240 245 250 255	
AAG GAT GAG GTT GAC ATC GTT GAG AAT GTC TAT GTG TAC AAC ATC TCT 1685	
Lys Asp Glu Val Asp Ile Val Glu Asn Val Tyr Val Tyr Asn Ile Ser	
260 265 270	
ATG TTT AAT GCT TCG GTA TGT CTG AAC TTT AAC CAT ATA ATA GAC TTC 1733	
Met Phe Asn Ala Ser Val Cys Ile Asn Phe Asn His Ile Ile Asp Phe	
275 280 285	
TTA CTA ACT TGG TTG CAG GAT ATG GCC CGC ATC AAG GTT TGG CCT GGT 1781	
Leu Leu Thr Trp Leu Gln Asp Met Ala Arg Ile Lys Val Trp Pro Gly	
290 295 300	
ACT CCC TCT GCG CTA TCT GCC GAT CTT CAA GGC GGC GGT GGC TAG GGT 1829	
Thr Pro Ser Ala Leu Ser Ala Asp Leu Gln Gly Gly Gly Ser Gly	
305 310 315	
AGC GTA AAG AAT ATC ACC TAT GAC ACC GCA CTC ATT GAT AAT GTC GAC 1877	
Ser Val Lys Asn Ile Thr Tyr Asp Thr Ala Leu Ile Asp Asn Val Asp	
320 325 330 335	
TGG GCC ATT GAA ATC ACG CAG TGC TAT GGG CAG AAG AAT ACT ACC TTG 1925	
Trp Ala Ile Glu Ile Thr Gln Cys Tyr Gly Gln Lys Asn Thr Thr Leu	
340 345 350	
TGC AAC GAG TAC CCG GTTAGTAGAC CTTCAGCCGC TTTCCCGAAG CTATCCTAAT 1980	
Cys Asn Glu Tyr Pro	
355	
ACAATAATAG AGC TCT CTC ACC ATC TCG GAC GTC CAC ATC AAG AAC TTC 2029	
Ser Ser Leu Thr Ile Ser Asp Val His Ile Lys Asn Phe	
360 365	
CGC GGT ACG ACG TCG GGA TCG GAA GAT CCC TAT GTT GGG ACA ATT GTT 2077	
Arg Gly Thr Thr Ser Gly Ser Glu Asp Pro Tyr Val Gly Thr Ile Val	
370 375 380 385	
TGT TCC AGT CCT GAT GTAAAGTGCCC TCCAGGATAT GCGTTAGTG TGCAATGGCT 2132	
Cys Ser Ser Pro Asp	
390	

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GACACTCGAT AG ACT TGC TCG GAT ATC TAT ACT TCC AAT ATT AAT GTA Thr Cys Ser Asp Ile Tyr Thr Ser Asn Ile Asn Val 395 400	2180
ACA AGC CCG GAT GGA ACC AAC GAC TTT GTT TGC GAT AAT GTGAGTOGNC Thr Ser Pro Asp Gly Thr Asn Asp Phe Val Cys Asp Asn 405 410 415	2229
CAAGGCCAGG TGRAGATCTA AATCGGTGAC TAACGGCTGTC CAG GTC GAT GAG AGT Val Asp Glu Ser	2284
CIT CIG AGT GTC AAC TGC ACC GCC ACT TCT GAT TAAGTTAGAA TGCCCTGTTAA Leu Leu Ser Val Asn Cys Thr Ala Thr Ser Asp 420 425 430	2337
CATGTACACT GGAGGTCAGG CCTTACCTAG ACGGATGAG CGGTTGATATA TTCACTTTGG GTTCCGGCTGT GTATATACGT GAGCAAATAA TATCTGTATG CTTTTGACIT AGGGAGGTC AGGCGATCAA GTAACCTTCT GGTCAAAACC TTGATCATGG TTCTCAGTAG ACGCTCCCTC TGGTTTTGCG GTTGGGATAC GTCACATGCC GAGGGAGGAA TCATTGCTTC CAGGTTGCTTG CGCTCTCCTG AGTGATACTG CACITGGCTGT CCAGGTTACT ACATCTCTCT GCATTACCT GCTGTTCATC AAACAAGCAA GCAACCTGAA GAGCCACCTA CCAAGTATAG TAGGATCTAG AGGGCTCCGA GGAGATGTTG TAGACCATAC TAGATCCTCG AAAATGTAAA TTGCAAACCA TATTTCATT CTAATTTCATC TGAAGTCCTGG CAGGGAGATA TATAACCGAA GTCTCTTGGAA ACACGAACCC TCGGCTGATG ATCGTGTACCT CCTGAGTAGC ACCCAACCCAG CGGACTACAA GATGGACAAC GAACCTTACC CACAACCTT AAGTACTTTG GAACCAAGTGTG GCAATGATTT GTATTCTTTA TCGGTGTGGG TCATGCACCT CCTGCAG	2397 2457 2517 2577 2637 2697 2757 2817 2877 2937 2974

(2) INFORMATION FOR SEQ ID NO: 5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 452 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

Met Arg Leu Thr His Val Leu Ser His Thr Leu Gly Leu Leu Ala Leu -22 -20 -15 -10
--

Gly Ala Thr Ala Glu Ala Phe Ser Arg Ser Arg Glu Ala Ala Cys Gly -5 1 5 10
--

Pro Lys Lys Pro Phe Arg Pro Leu Pro Thr Ser Gln Ser Arg Asp Lys

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15	20	25	
Thr Cys His Val Arg Ser His Gly Asp Gly	Thr Asp Asp Ser Asp Tyr		
30	35	40	
Ile Leu Ser Ala Leu Asn Gln Cys Asn His Gly	Gly Lys Val Val Phe		
45	50	55	
Asp Glu Asp Lys Glu Tyr Ile Ile Gly	Thr Ala Leu Asn Met Thr Phe		
60	65	70	
Leu Lys Asn Ile Asp Leu Glu Val Leu Gly	Thr Ile Leu Phe Thr Asn		
75	80	85	90
Asp Thr Asp Tyr Trp Gln Ala Asn Ser Phe	Lys Gln Gly Phe Gln Asn		
95	100	105	
Ala Thr Thr Phe Phe Gln Leu Gly	Gly Glu Asp Val Asn Met Tyr Gly		
110	115	120	
Gly Gly Thr Ile Asn Gly Asn Gly	Gln Val Trp Tyr Asp Leu Tyr Ala		
125	130	135	
Glu Asp Asp Leu Ile Leu Arg Pro	Ile Leu Met Gly Ile Ile Gly Leu		
140	145	150	
Asn Gly Gly Thr Ile Gly Pro Leu Lys	Leu Arg Tyr Ser Pro Gln Tyr		
155	160	165	170
Tyr His Phe Val Ala Asn Ser Ser Asn	Val Leu Phe Asp Gly Ile Asp		
175	180	185	
Ile Ser Gly Tyr Ser Lys Ser Asp Asn	Glu Ala Lys Asn Thr Asp Gly		
190	195	200	
Trp Asp Thr Tyr Arg Ser Asn Asn	Ile Val Ile Gln Asn Ser Val Ile		
205	210	215	
Asn Asn Gly Asp Asp Cys Val Ser Phe	Lys Pro Asn Ser Thr Asn Ile		
220	225	230	
Leu Val Gln Asn Leu His Cys Asn Gly	Ser His Gly Ile Ser Val Gly		
235	240	245	250
Ser Leu Gly Gln Tyr Lys Asp Glu Val	Asp Ile Val Glu Asn Val Tyr		
255	260	265	
Val Tyr Asn Ile Ser Met Phe Asn Ala	Ser Val Cys Leu Asn Phe Asn		
270	275	280	
His Ile Ile Asp Phe Leu Leu Thr Trp	Leu Gln Asp Met Ala Arg Ile		
285	290	295	
Lys Val Trp Pro Gly Thr Pro Ser Ala	Leu Ser Ala Asp Leu Gln Gly		
300	305	310	
Gly Gly Gly Ser Gly Ser Val Lys Asn	Ile Thr Tyr Asp Thr Ala Leu		
315	320	325	330

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Ile Asp Asn Val Asp Trp Ala Ile Glu Ile Thr Gln Cys Tyr Gly Gln
 335 340 345
 Lys Asn Thr Thr Leu Cys Asn Glu Tyr Pro Ser Ser Leu Thr Ile Ser
 350 355 360
 Asp Val His Ile Lys Asn Phe Arg Gly Thr Thr Ser Gly Ser Glu Asp
 365 370 375
 Pro Tyr Val Gly Thr Ile Val Cys Ser Ser Pro Asp Thr Cys Ser Asp
 380 385 390
 Ile Tyr Thr Ser Asn Ile Asn Val Thr Ser Pro Asp Gly Thr Asn Asp
 395 400 405 410
 Phe Val Cys Asp Asn Val Asp Glu Ser Leu Leu Ser Val Asn Cys Thr
 415 420 425
 Ala Thr Ser Asp
 430

(2) INFORMATION FOR SEQ ID NO: 6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 17 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iii) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: pki SmaI sense

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

TTTTCCCTTCC CGGGCAC

17

(2) INFORMATION FOR SEQ ID NO: 7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 30 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)

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- (iii) HYPOTHETICAL: NO
- (iii) ANTI-SENSE: YES
- (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: pki-pgaX antisense

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

GTGGCTGAGT CTCATCTTGA CGGATGATG

30

- (2) INFORMATION FOR SEQ ID NO: 8:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 30 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iii) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: pki-pgaX sense

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

CAATCATCCG TCAAGATGAG ACTCACGGAC

30

- (2) INFORMATION FOR SEQ ID NO: 9:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 17 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iii) ANTI-SENSE: YES
- (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: pgaX NcoI antisense

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

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AGTGCCATCT CCATGGC

17

(2) INFORMATION FOR SEQ ID NO: 10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 23 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: YES

(vi) ORIGINAL SOURCE:

- (C) INDIVIDUAL ISOLATE: 3062

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: one-of(3, 6, 9, 15)
- (D) OTHER INFORMATION: /note= "N at position 3, 6, 9, and 15 is inosine"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

GGNGTNCCNG GVCANACYTT DAT

23

(2) INFORMATION FOR SEQ ID NO: 11:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 23 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

- (C) INDIVIDUAL ISOLATE: 3063

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: one-of(9, 15, 18)
- (D) OTHER INFORMATION: /note= "N at position 9, 15, and 18 is inosine"

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:

ATTAARGINT GBCCNGGNAC NCC

23

(2) INFORMATION FOR SEQ ID NO: 12:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 32 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: YES

(vi) ORIGINAL SOURCE:

- (C) INDIVIDUAL ISOLATE: HR7734

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: one-of(3, 9, 12, 15, 21, 23)
- (D) OTHER INFORMATION: /note= "N at position 3, 9, 12, 15, 21, and 23 is inosine"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

GGNCCDATNG TNCCNCCRIT NARNCCDATD AT

32

(2) INFORMATION FOR SEQ ID NO: 13:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 29 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

- (C) INDIVIDUAL ISOLATE: 3059

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: one-of(9, 12, 18, 21, 24)
- (D) OTHER INFORMATION: /note= "N at position 9, 12, 18, 21, and 24 is inosine"

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:

ATHATHGGNY TNAAYGGNGG NACNATHGG

29

(2) INFORMATION FOR SEQ ID NO: 14:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 23 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

- (C) INDIVIDUAL ISOLATE: 3060

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: one-of(6, 9, 12, 15)
- (D) OTHER INFORMATION: /note= "N at position 6, 9, 12, and 15 is inosine"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

GAGGCNGCNA CNGGNCCNAA GAA

23

(2) INFORMATION FOR SEQ ID NO: 15:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 23 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: YES

(vi) ORIGINAL SOURCE:

- (C) INDIVIDUAL ISOLATE: 3061

(ix) FEATURE:

- (A) NAME/KEY: misc_feature
- (B) LOCATION: one-of(6, 9, 15)
- (D) OTHER INFORMATION: /note= "N at position 6, 9, and 15

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is inosine"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 15:

TTTTTNGGNC CTTTNGCNGC YTC

23

Claims

1. A purified and isolated DNA sequence which encodes a polypeptide having the activity of the Aspergillus tubigensis PGX enzyme, characterized in that the DNA sequence comprises,
 - a) the sequence of SEQ ID No.1,
 - b) genetic variants of the sequence of a),
 - c) a DNA sequence capable of hybridizing with the sequences of a) or b) or with parts thereof.
- 10 2. A DNA construct containing a DNA sequence according to claim 1, operably linked to regulatory regions capable of directing the expression of said DNA sequence in a suitable host organism.
- 15 3. A DNA vector containing the construct of claim 2.
4. A transformed microbial host cell capable of expressing a gene encoding a protein having exo-polygalacturonase activity characterized in that the host cell contains a DNA vector according to claim 3.
- 20 5. A method for obtaining expression of exo-polygalacturonase comprising, culturing the microbial host cell of claim 4 under conditions wherein the cloned gene is expressed.
- 25 6. A method for obtaining exo-polygalacturonase comprising the steps of
 - a) culturing the microbial host cell of claim 4 under conditions giving rise to the expression of the DNA sequence of claim 1,
 - b) recovering the polypeptide having PGX activity.
- 30 7. A polypeptide having PGX activity characterized in that the polypeptide is produced by the method of claim 6.

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8. Use of a polypeptide obtained according to claim 7 and having exo-polygalacturonase activity to degrade a polymer containing glucuronic acid containing residues.
9. Purified and isolated expression and transcription regulatory regions as found in the 5'non-coding sequences of the Aspergillus tubiqensis pqaX gene.
10. pIM362

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1/4

FIGURE 1

a:

5 5' G G I G T I C C I G G C C A I A C T T A A T 3' (3062) (non-coding strand)
 A C T
 G G

10

5' A T T A A G T I T G G C C I G G I A C G C C 3' (3063) (coding strand)
 C G T A
 A C C T

15

b:

20

5' G G I C C A A T I G T I C C I C C A T T I A A I C C A A T A A T 3' (7734) (non-coding
 T T
 G G G G strand)

25

30

5' A T T A T T G G I T T I A A T G G I G G I A C I A T T G G I C C 3' (3059) (coding
 A
 C C C C strand)

c:

35

5' G A A G C I G C I A A I G G I C C C A A G A A 3' (3060) (coding
 A A
 G C G T strand)

40

45

5' T T C T T I G G I C C G T T I G C I G C T C 3' (3061) (non-coding
 T A C strand)

50

a: derived from CB 1, residues 3-10

b: derived from CB 2, residues 2-12

c: derived from CB 3, residues 6-13

55

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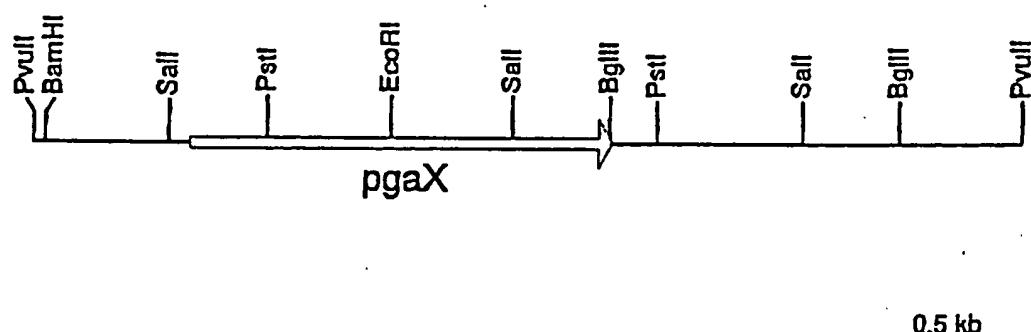


Figure 2

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FIGURE 3

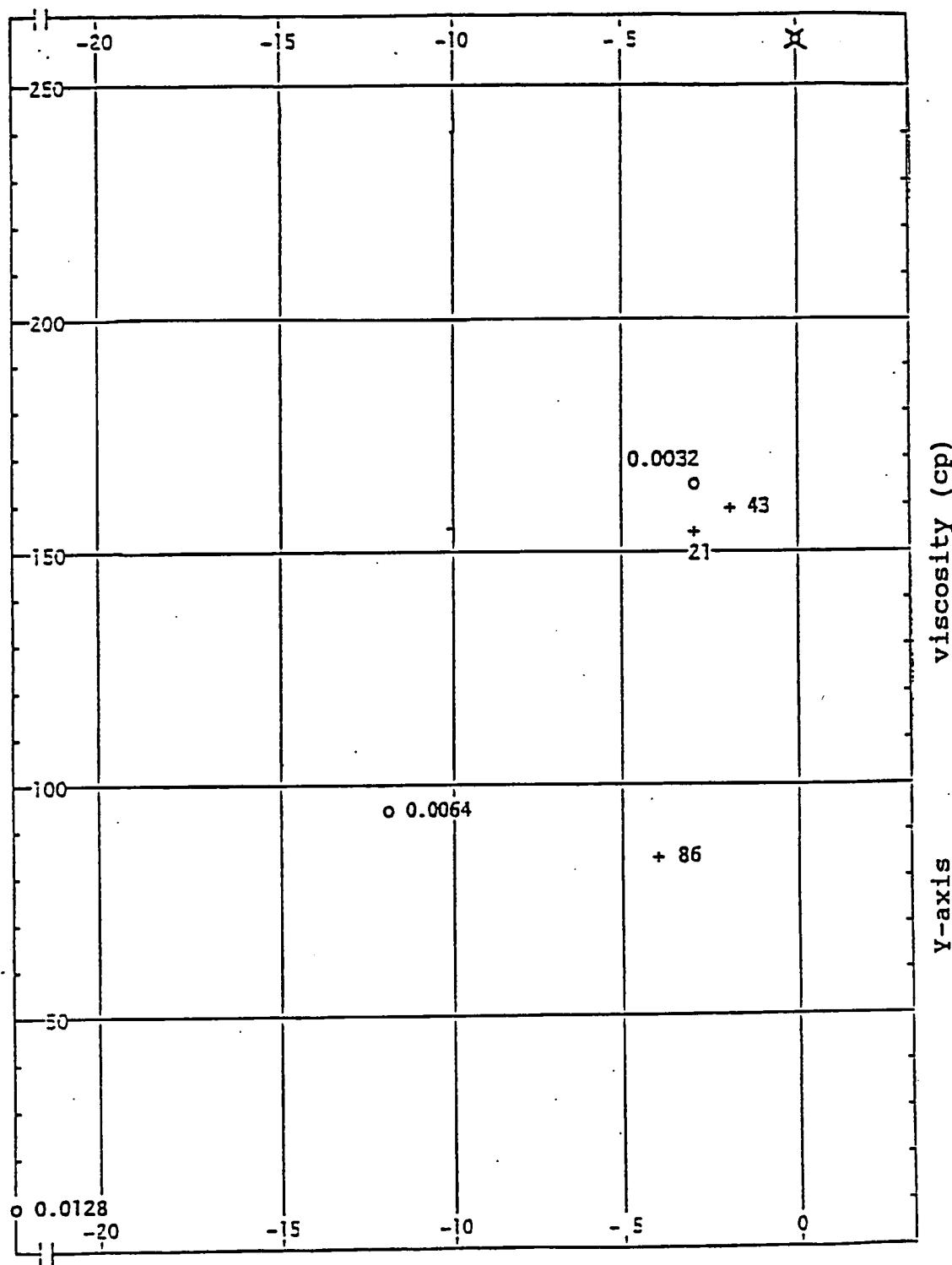
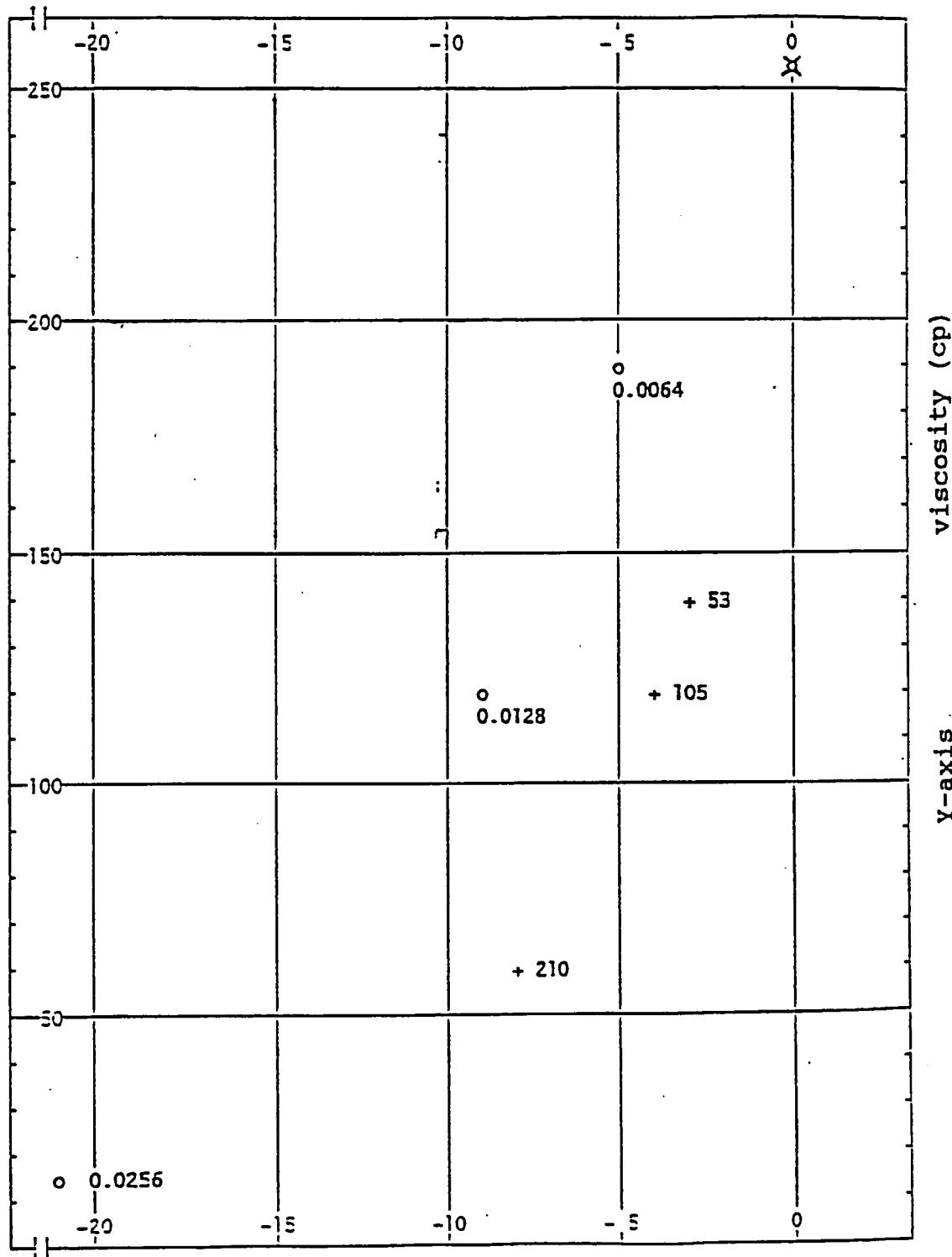
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X-axis change in MW (kD)

FIGURE 4

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X-axis change in MW (kD)



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INTERNATIONAL SEARCH REPORT

International Application No

PCT/EP 93/03704

A. CLASSIFICATION OF SUBJECT MATTER
 IPC 5 C12N15/56 C12N15/11 C12N9/24

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
 IPC 5 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	EP,A,0 421 919 (CYBA-GEIGY AG) 10 April 1991 cited in the application see page 4, line 2 - line 14 see page 5, line 12 - line 33 see page 5, line 43 - page 6, line 15 see page 8, line 17 - line 21 --- WO,A,91 04331 (GENESIT OY) 4 April 1991 see page 4, line 30 - page 5, line 12 see page 10, line 7 - page 11, line 29 --- -/--	1-10
A		1-6

Further documents are listed in the continuation of box C.

Patent family members are listed in annex.

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Date of the actual completion of the international search

16 March 1994

Date of mailing of the international search report

25. 94 94

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INTERNATIONAL SEARCH REPORT

Inte nal Application No
PCT/EP 93/03704

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>CURRENT GENETICS vol. 19, no. 6, 1991 pages 467 - 474 HENDRIK J.D. BUSSINK ET AL. 'Expression and sequence comparison of the <i>Aspergillus</i> <i>niger</i> and <i>Aspergillus</i> <i>tubigensis</i> genes encoding polygalacturonase II' see page 467, right column, last paragraph see page 468, right column, paragraph 1 see page 469, left column, paragraph 3; figure 5 -----</p>	1-5

INTERNATIONAL SEARCH REPORT

Information on patent family members

Int: **na** Application No
PCT/EP 93/03704

Patent document cited in search report	Publication date	Patent family member(s)		Publication date
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		AU-A-	6205490	07-03-91
		JP-A-	3224489	03-10-91
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		EP-A-	0491743	01-07-92
		JP-T-	5500309	28-01-93
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